Mini focus on bioresorbable scaffolds

Preclinical investigation of neoatherosclerosis in magnesiumbased bioresorbable scaffolds versus thick-strut drug-eluting stents



Philipp Nicol¹, MD; Anna Bulin¹, DVM, MD; Maria Isabel Castellanos^{1,2}, PhD; Magdalena Stöger¹, MS; Simone Obermeier¹, MS; Jonas Lewerich¹, MS; Tobias Lenz¹, MD; Petra Hoppmann^{2,3}, MD; Christine Baumgartner⁴, DVM; Johannes Fischer⁴, DVM; Katja Steiger⁵, DVM; Michael Haude⁶, MD; Michael Joner^{1,2*}, MD

1. Deutsches Herzzentrum München and Deutsches Zentrum für Herz-Kreislaufforschung e.V., Munich, Germany; 2. DZHK (German Centre for Cardiovascular Research), partner site Munich Heart Alliance, Munich, Germany; 3. Klinik und Poliklinik für Innere Medizin I, Klinikum rechts der Isar, Technische Universität München, Munich, Germany; 4. Zentrum für präklinische Forschung, Technische Universität München, Munich, Germany; 5. Institut für Pathologie, Technische Universität München, Munich, Germany; 6. Städtische Kliniken Neuss, Lukaskrankenhaus GmbH, Neuss, Germany

P. Nicol and A. Bulin contributed equally to this manuscript.

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KEYWORDS

- bioresorbable scaffolds
- drug-eluting stent • optical coherence
- tomography

Abstract

Aims: Neoatherosclerosis is a frequent finding after implantation of permanent metallic stents. Bioresorbable scaffolds (BRS) are considered to reduce the incidence of neoatherosclerosis owing to their dissolution and consequent vascular restoration. The aim of this study was to evaluate the formation of neoatherosclerosis between magnesium-based BRS and thick-strut metallic drug-eluting stents (DES) in a rabbit model of neoatherosclerosis and in proportion to the effect of high-dose statin medication.

Methods and results: Fully bioresorbable magnesium scaffolds (BRS, n=45) and thick-strut permanent metallic DES of equivalent geometry and design (n=45) were implanted into the iliac arteries of New Zealand White rabbits (n=45) following endothelial balloon injury and exposure to a cholesterol diet. Endothelialisation was assessed in 12 animals after 35 days using scanning electron microscopy (SEM), showing significantly enhanced re-endothelialisation above struts in the BRS (n=13) compared to DES (n=10). Eleven (11) animals were terminated for baseline assessment after 91 days while the remaining 22 animals were randomised to receive high-dose statin treatment (3 mg/kg) or placebo. BRS-treated vessels showed a significant reduction in foam cell infiltration as a sign of early neoatherosclerosis by histology and OCT when compared to thick-strut DES-treated vessels. Statin treatment resulted in significant reduction in BRS and DES by histology.

Conclusions: Our findings suggest reduced neoatherosclerosis formation in magnesium-based BRS relative to thick-strut DES. High-dose statin treatment may be a promising measure to reduce neoatherosclerosis progression, both on its own and in synergy with site-targeted device-based treatment.

*Corresponding author: German Heart Center Munich, Lazarettstrasse 36, 80636 Munich, Germany. E-mail: joner@dhm.mhn.de

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Abbreviations

atm	atmosphere
BRS	bioresorbable scaffold(s)
CI	confidence interval
DAH	delayed arterial healing
DES	drug-eluting stent
GEE	generalised estimating equation(s)
MMA	methyl methacrylate
OCT	optical coherence tomography
PLLA	poly-L-lactic acid
SEM	scanning electron microscopy

Introduction

Atherothrombotic cardiovascular disease is the leading cause of morbidity and mortality worldwide¹. In patients with obstructive coronary artery disease, percutaneous angioplasty and stenting is the first treatment of choice². However, while a considerable reduction of in-stent restenosis was achieved with the introduction of drug-eluting stents (DES), the undoubted success of this technology is delivered at the collateral cost of delayed arterial healing (DAH), a pathophysiological entity which underlies a spectrum of adverse clinical events^{3,4}. In-stent neoatherosclerosis has recently been recognised as an important disease entity in patients after stent implantation, representing an additional manifestation of atherosclerotic disease found in nascent neointimal tissue after stent implantation⁴.

Bioresorbable scaffolds (BRS) have recently been introduced into clinical practice. It has been proposed that their transient presence in the coronary artery may mitigate neoatherosclerosis formation. While first-generation polymer-based BRS are hampered by inherent shortcomings such as excessive strut thickness and slow degradation profile⁵, newer device iterations are targeted to address these limitations and provide a more rapid vascular restoration⁶. Furthermore, pharmacological targeting of risk factors such as hyperlipidaemia using high-dose statin treatment significantly decreased mortality in secondary prevention after manifestation of coronary artery disease⁷. Consequently, we aimed to investigate whether site-targeted treatment using magnesium-based BRS technology mitigates neoatherosclerosis formation in a novel hypercholesterolaemic animal model when compared to local control therapy with permanent metallic DES. It was another goal of this study to investigate the effect of BRS versus DES progression of neoatherosclerosis in proportion to systemic treatment with atorvastatin compared to placebo. The study flow is shown in **Figure 1**.

Methods

Please refer to **Supplementary Appendix 1** for details regarding materials and methods.

Results

Of the 33 rabbits included in the histology study, two animals died of cholesterol-induced liver failure. Two additional animals had to be excluded from analysis due to total thrombotic occlusion of a stented vessel (one rabbit with thrombosis in BRS, one with thrombosis in DES). Accordingly, a total of nine animals were available in group 1 (day 91). A total of 10 animals each remained in groups 2 and 3, respectively, until day 161.

Serum levels of cholesterol differed significantly between groups with an area under the curve (AUC) of 3608.97 in group 1 (baseline) and an AUC of 3319.28 in group 2 (statin treatment) versus 4016.92 in group 3 (placebo treatment; p=0.02) (Supplementary Figure 1).

The results of group 1 are shown in **Supplementary Table 1**, **Supplementary Table 2**, and **Supplementary Appendix 2**. There was no early mortality in the 12 rabbits assigned to scanning electron microscopy (SEM) assessment of endothelialisation.



Figure 1. Study flow chart. All animals received BRS and DES in the iliac arteries seven days after the start of a 1% cholesterol diet and were switched to a low-dose cholesterol diet on day 35. Repeat balloon denudation was performed on day 63. On day 91, animals of group 1 were terminated while the remaining animals were randomised to statin treatment or placebo. The animals in groups 2 and 3 were euthanised on day 161.

NEOATHEROSCLEROSIS SCORING BY HISTOLOGY

BRS showed a significantly lower estimated score of foamy macrophage infiltration compared to DES (score 1.16 [1.12, 1.21] vs 1.53 [1.41, 1.65]; mean reduction -0.361; p<0.0001) (Figure 2A). A neointimal foam cell score was additionally assigned to each quadrant of histological cross-sections, treated as a categorical variable and compared among treatment groups (BRS vs DES). Each quadrant showed significant differences based on Pearson's chi square, with a higher percentage of foam cell score 2-4 in DES relative to BRS (p<0.05 for all quadrants).

Statin treatment resulted in a significant reduction of foamy macrophage infiltration relative to placebo (score 1.23 [1.14, 1.32] vs 1.45 [1.36, 1.54]; mean reduction -0.221; p=0.001) (Figure 2B, Table 1). Variability of scoring was assessed by calculating the intraclass and interclass correlation coefficient (0.98 and 0.90, respectively).

There was significant statistical interaction (p=0.005) among stent types and treatment allocation to statin versus placebo. Figure 3 shows representative co-registration of haematoxylin



Figure 2. Neoatherosclerosis in histological sections (n=20 rabbits). Estimated mean neoatherosclerosis score derived by generalised estimating equations (lower/upper CI) shows a significant reduction of foamy macrophage infiltration in BRS as compared to DES (A) and statin-treated animals versus placebo (B).

and eosin (H&E)-stained histological slides and optical coherence tomography (OCT) frames in both BRS and DES.



Figure 3. Comparison and co-registration of BRS and DES. A) Sections stained for H&E (A1) with corresponding OCT frame (A2) in a BRS-treated vessel. B) Sections stained for H&E (B1) with corresponding OCT frame (B2) in a DES-treated vessel. Asterisks (*) indicate foamy macrophage infiltration.

NEOATHEROSCLEROSIS SCORING BY OCT

An overview of the results is presented in **Table 2**. BRS showed a significantly lower estimated score of macrophage infiltration by OCT compared to DES (score 1.20 [1.08, 1.34] vs 2.09 [1.83, 2.39]; mean reduction -0.886; p<0.0001) (Figure 4A). There was no statistically significant difference in animals receiving statin treatment relative to placebo by OCT (score 1.57 [1.32, 1.87] vs 1.60 [1.44, 1.77]; mean reduction -0.26; p=0.873) (Figure 4B). Variability of scoring was assessed by calculating the intraclass and interclass correlation coefficient (0.74 and 0.69, respectively).

MORPHOMETRIC ANALYSIS IN OCT AND HISTOPATHOLOGY

Morphometric assessment revealed significantly greater neointimal area in BRS relative to DES by OCT (1.86 mm² vs 1.32 mm²; mean difference +0.541 in BRS; p<0.0001) and histology (3.22 mm²)

	•	•				•		-			
			Neoath	erosclerosi	s score		% Stenosis				
		Mean	Lower 95% Cl	Upper 95% Cl	<i>p</i> -value	B-value	Mean	Lower 95% Cl	Upper 95% Cl	<i>p</i> -value	B-value
Treatment	BRS, n=10	1.16	1.12	1.21	0.0001*	0.201	68.64	64.82	72.69	0.0001*	0.670
	DES, n=10	1.53	1.41	1.65	0.0001	-0.391	35.53	32.33	39.04	0.0001	0.079
Group	Statin, n=10	1.23	1.14	1.32	0.001*	1* 0.000	49.33	44.22	55.03	0.077	0.019
	Placebo, n=10	1.45	1.36	1.54	0.001	-0.266	49.43	45.39	53.84	0.977	0.018
p for intera	ction		0.0	05*				0.5	577		
Values are presented as estimated means with upper and lower 95% Cl. A <i>p</i> <0.05 was considered statistically significant (*). Wilcoxon Kruskal-Wallis (non-parametric) or ANOVA (parametric) was applied.											



Figure 4. Neoatherosclerosis in OCT (n=20 rabbits). Estimated mean neoatherosclerosis score derived by generalised estimating equations (lower/upper CI) assessed in OCT imaging shows a significant reduction of foamy macrophage infiltration in BRS as compared to DES (A) and a non-significant reduction in the statin-treated versus the placebo group (B).

vs 2.27 mm²; mean difference +0.948 in BRS; p<0.0001). This resulted in increased percentage stenosis in BRS-treated arteries relative to DES – 47% in BRS vs 21.8% in DES (p<0.0001) in OCT analysis (**Table 2**) and 68.6% vs 35.5% (p<0.0001) in histological analysis (**Table 1**, Figure 5).

HISTOLOGICAL AND IMMUNOHISTOLOGICAL STAINING

Neointimal tissue consisted of smooth muscle cells in a proteoglycan-rich matrix. Inflammation of neointimal tissue, consisting of lymphocytes and for the largest part neutrophilic granulocytes, was generally mild to moderate with a tendency towards greater inflammation in BRS relative to DES (inflammation score: 1.53 in BRS vs 1.24 in DES, p=0.106). Cholesterol crystals were frequently observed in deeper tissue layers in both BRS and DES. Giant cells were mainly observed within the peri-strut regions (% giant cells: 9.33% in BRS vs 7.70% in DES, p=0.335). In segments stained for RAM11, macrophage infiltration was frequently



Figure 5. Percentage stenosis comparing BRS versus DES in OCT and histopathology (n=20 rabbits). A) Estimated mean percentage stenosis derived by generalised estimating equations (lower/upper CI) as assessed by morphometrical analysis of histology in both BRS and DES. B) Estimated mean percentage stenosis derived by generalised estimating equations (lower/upper CI) as assessed by OCT imaging in both BRS and DES.

observed within the subintimal tissue in DES, while BRS showed scattered macrophages within the peri-strut tissue (Figure 6). Additionally, staining against smooth muscle actin (SMA) showed formation of mature smooth muscle cells in BRS (Figure 7A1, Figure 7A2), while DES did not show any positive staining for SMA (Figure 7B1, Figure 7B2). For detailed results of histopathological assessment and morphometry in histology and OCT please refer to Supplementary Table 3 and Supplementary Table 4.

ENDOTHELIAL CELL COVERAGE

Re-endothelialisation following catheter denudation and stenting was evaluated in 12 animals (n=13 BRS, n=10 DES) using SEM. Twenty-eight days following stent implantation (day 35), magne-sium-based BRS showed significantly improved endothelial cell coverage above struts as compared to the 316L DES of equivalent design (91.7 \pm 20.2% vs 42.1 \pm 57.16%; p<0.001) (Figure 8).

Table 2. Results of OCT	analysis of groups 2 and	3 after 161 days (n=10 rab	bits for each group).
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			•				0				
			Neoath	erosclerosi	s score		% Stenosis				
		Mean	Lower 95% Cl	Upper 95% Cl	<i>p</i> -value	B-value	Mean	Lower 95% Cl	Upper 95% Cl	<i>p</i> -value	B-value
Treatment	BRS, n=10	1.20	1.08	1.34	0.0001*	0.606	47.00	41.44	53.32	0.0001*	0.975
	DES, n=10	2.09	1.83	2.39	0.0001^	-0.606	21.77	18.82	25.20	0.0001	0.675
Group	Statin, n=10	1.57	1.32	1.87	0 072	0.070	32.31	27.03	38.61	0.857	0.125
	Placebo, n=10	1.60	1.44	1.77	- 0.873 -0.070 31.68 28.14 35.67 0.8		0.857	0.125			
p for intera	ction		0.4	35			0.222				
Values are presented as estimated means with upper and lower 95% CI. A p<0.05 was considered statistically significant (*). Wilcoxon Kruskal-Wallis (non-parametric) or ANOVA (parametric) was applied.										al-Wallis	



Figure 6. *RAM-11 staining in BRS and DES stented vessels. A) Overview of a RAM-11 stained section from a BRS (A1) with 40x magnification (A2). Sparse macrophage infiltration is detectable in peri-strut neointimal tissue. B) Overview of a RAM-11 stained section from a DES (B1), with 40x magnification (B2). Superficial foamy macrophage infiltration is visible in about 50% of the vessel circumference.*

Discussion

The current study investigated the differential magnitude of neoatherosclerosis formation after implantation of fully bioresorbable magnesium scaffolds and permanent metallic thick-strut DES of equivalent design and geometry in a novel animal model of instent neoatherosclerosis. Furthermore, it examined the impediment of neoatherosclerosis progression as a response to statin treatment relative to placebo in both stent types up to 161 days following study initiation. The most salient findings can be described as follows:

(i) Fully bioresorbable magnesium scaffolds showed enhanced endothelial cell coverage 28 days following stent implantation and

decreased neointimal macrophage infiltration after 161 days when compared to permanent stainless steel thick-strut DES in a novel animal model of neoatherosclerosis.

(ii) There was significant statistical interaction between stent type and treatment allocation to atorvastatin versus placebo with regard to reduction of neointimal macrophage infiltration. Therefore, synergistic effects between local and systemic therapy are very likely.
(iii) Atorvastatin treatment as applied in the current study significantly reduced progression of neointimal macrophage infiltration in BRS and DES, which was confirmed by histopathological assessment.
(iv) Fully bioresorbable magnesium scaffolds showed significantly greater neointima burden as compared to thick-strut DES.

Development of neoatherosclerosis after stent implantation has been recognised as an important contributor to late stent failure. The implantation of DES was recently reported to result in earlier onset of neoatherosclerosis formation in post mortem autopsy and clinical imaging studies when compared to BMS^{8,9}. Previous work has shown that DES exhibit markedly different neointimal composition with a higher amount of proteoglycan and fewer smooth muscle cells compared with BMS^{10,11}. Notably, long-term follow-up of clinical trials investigating the comparative performance of earlyand newer-generation DES reported a sustained increase in clinical events irrespective of the implanted stent type over time¹². Short of a mechanistic explanation for this late increment in adverse events, preclinical and post-mortem autopsy studies have suggested delayed arterial healing with sustained inflammatory reaction, delayed endothelialisation and formation of in-stent neoatherosclerosis to be causative in the majority of cases. Consequently, bioresorbable scaffolds were introduced to overcome these inherent limitations associated with permanent metallic DES and proposed to reduce the burden of neoatherosclerosis owing to their temporary presence, where recovery of physiological vasomotion and normalisation of flow dynamics were suggested to impact favourably on plaque progression over time. However, those anticipated effects of BRS technology have not been proven in clinical practice.



Figure 7. *SMA staining in BRS and DES stented vessel. H&E stained sections in BRS (A1) and DES (B1) show the presence of spindle-shaped cells, indicating smooth muscle cells (40x magnification in A2/B2). A3) & A4) SMA stained section from a BRS with 40x magnification. Positive areas reveal these cells to be mature smooth muscle cells (arrows). B3) & B4) SMA stained section from a DES. No positive areas are visible, indicating the absence of mature smooth muscle cells.*



Figure 8. Endothelial cell coverage in BRS and DES. Left: scanning electron microscopy of BRS (upper panel) and DES (lower panel) with higher magnification of boxed areas. Right: quantification of endothelial cells above stent struts.

Since reliable assessment of neoatherosclerosis by OCT alone remains controversial and prospective invasive imaging studies are challenging to perform, preclinical studies remain an important means to investigate the comparative effects of medical devices and pharmacological approaches in a timely manner. We have previously validated the currently applied animal model and found excellent agreement with regard to depiction of early signs of neoatherosclerosis such as foam cell infiltration within the nascent neointima following stent implantation¹³. Since early stages of neoatherosclerosis can be detected reliably by OCT (i.e., macrophage infiltration), the currently applied animal model holds great potential to gain a mechanistic understanding into the pathophysiology and adoption of therapeutic strategies for the treatment of neoatherosclerosis. The application of two distinct scoring systems for detecting neoatherosclerosis could have biased our results, especially when detecting neoatherosclerosis by OCT. However, reproducibility was satisfactory and variability rather low in both scores.

Owing to the inherent design constraints of the current animal study, absolute effect size in reducing neoatherosclerosis among BRS versus DES and atorvastatin versus placebo-treated animals is difficult to ascertain. This is also reflected by significant statistical interaction among these treatment approaches with regard to reduction of neointimal macrophages. It can therefore be assumed that BRS and atorvastatin therapy resulted in synergistic effects with regard to neoatherosclerosis progression over time, resulting in reduced neointimal macrophage infiltration.

Bioresorbable scaffold technology has recently suffered a major setback after the release of randomised controlled clinical trial data, which showed significantly increased revascularisation and device thrombosis rates at longer-term follow-up in broad patient subsets^{14,15}. Potential mechanisms of late scaffold failure have been investigated in dedicated registries using OCT evaluating the long-term outcomes of polymeric BRS to delineate vascular morphology. These suggested late strut discontinuities, malapposition, neoatherosclerosis and underexpansion of the scaffold segment to be causally associated¹⁶. While most of these findings can be attributed to scaffold dismantling over time, neoatherosclerosis is not specifically linked to bioresorbable scaffold technology, and was indeed forecast to be erased with the use of BRS.

In our study, we observed decreased foam cell infiltration after implantation of magnesium-based BRS relative to a thick-strut permanent metallic DES, which may be secondary to accelerated vascular restoration (within six months) after magnesium-based BRS implantation relative to the three-year time frame observed with first-generation polymeric BRS, return to physiologic flow conditions and accelerated recovery of functional endothelial barrier function. Drug-induced impaired re-endothelialisation is likely to play a key role in the development of neoatherosclerosis as it allows the passage of inflammatory cells and lipoproteins into the developing neointima¹⁷. Quantitative evaluation of re-endothelialisation after 28 days showed significantly improved endothelial cell coverage in BRS compared with thick-strut DES. However, the lack of sufficient evaluation of endothelial leakiness in our study is a limitation. Our data are also in contrast to previous work by Waksman showing a lower degree of endothelialisation 28 days following stent implantation¹⁸: the main difference to our study can be seen in the procedural methodology of balloon denudation and the usage of healthy versus atherosclerotic animals. Most importantly, re-endothelialisation following arterial denudation is variable and depends largely on animal species, diseased versus healthy vascular tissue conditions, and the degree of vascular injury performed prior to stent implantation. In the study by Waksman et al, standard semicompliant balloon catheters were inflated at the target site of stent implantation in healthy rabbits to achieve endothelial denudation.

In the current study, we used 3 Fr Fogarty catheters, which were inflated and repeatedly pulled back (arterial embolectomy catheters) to cause endothelial denudation in atherosclerotic rabbits. In our experience, the latter methodology causes a greater degree of endothelial denudation and vascular injury. As a consequence of greater endothelial injury in our study, proliferation and migration of endothelial cells is also markedly increased, which may explain the enhanced rate of re-endothelialisation 28 days after stent implantation. However, it is important to mention that re-endothelialisation was assessed at day 35 in this animal model, a time point at which lacking endothelial integrity is likely to impact substantially on cholesterol uptake into neointimal tissue and, consequently, formation of foamy macrophages. The fact that we observed increased foamy macrophage infiltration in thick-strut DES relative to BRS further strengthens this hypothesis. Atorvastatin has previously been proven to result in decreased plaque formation in hypercholesterolaemic rabbits^{19,20}. There is abundant evidence of its clinical efficacy from large randomised controlled trials7. Atorvastatin was found to result in a highly significant 72% decrease of iliac-femoral lesion size following balloon injury in hypercholesterolaemic rabbits, which was attributed mainly to a significant reduction of intimal macrophages²⁰. In a similar vein, Herdeg et al found a significant reduction in intimal and medial proliferation as well as peristrut inflammation after atorvastatin therapy and bare metal stent implantation¹⁹, which is in line with the findings of our study, where atorvastatin therapy resulted in significantly decreased neointimal macrophage infiltration irrespective of stent type.

In our study, atorvastatin relative to placebo therapy resulted in significantly diminished progression of neointimal foam cell infiltration, which was independent from the underlying stent type.

Limitations

Firstly, animal models are never able to replicate fully all the features of certain pathologies, e.g., we only observed the early stages of neoatherosclerosis and no advanced stages such as necrotic core formation. Also, the formation of foam cells is artificially accelerated (e.g., repeat endothelial denudation) compared to human coronary atherosclerosis, which usually takes decades to develop and depends on additional important co-factors that cannot be reproduced in current animal models.

Secondly, OCT has limitations in the detection of neoatherosclerosis, as mentioned above. This can be challenging when using degrading stent struts as a hallmark to differentiate neointima from native intima in OCT and could have biased the results of neoatherosclerosis scoring. However, even after degradation, BRS struts could be reliably recognised in the majority of cases and helped to differentiate progression of atherosclerosis versus neoatherosclerosis. Additionally, the use of co-registered histology allowed reliable diagnosis of neoatherosclerosis. Thirdly, transferability of preclinical findings to a human disease state is difficult to ascertain even after applying diseased animal models.

Finally, the use of a thick-strut DES is a limitation since modern DES have much thinner struts; thus, comparison to contemporary

DES used in clinical practice with respect to neoatherosclerosis remains unknown.

Consequently, the current study should be viewed as hypothesisgenerating. Clinical trials designed specifically to address neoatherosclerosis formation over time are needed to confirm these findings.

Conclusion

This study is the first to suggest reduction of in-stent neoatherosclerosis in bioresorbable magnesium scaffolds compared to thickstrut DES. High-dose statin therapy may be an important secondary prevention measure in patients with confirmed neoatherosclerosis. Prospective randomised trials are needed to evaluate these preclinical findings.

Impact on daily practice

Neoatherosclerosis was reported to contribute substantially to late stent failure, mainly caused by in-stent restenosis and stent thrombosis beyond the first year after stent implantation. In the current study, we investigated a second-generation magnesiumbased BRS with regard to formation of neoatherosclerosis in hypercholesterolaemic rabbits relative to its stainless steel equivalent DES and found significantly reduced foam cell formation as an early sign of neoatherosclerosis in magnesiumbased BRS. Clinical studies with appropriately designed endpoints are required to confirm these promising results in man.

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Conflict of interest statement

M. Haude reports personal fees for consulting from Biotronik, outside the submitted work. M. Joner reports grants from the ESC's "Grants for Medical Research Innovation", during the conduct of the study, personal fees for consulting from Biotronik and OrbusNeich, and personal fees from speaker fees from Biotronik, OrbusNeich, Boston Scientific, Medtronic, and AstraZeneca, outside the submitted work. The other authors have no conflicts of interest to declare.

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Supplementary data

Supplementary Appendix 1. Methods.

Supplementary Appendix 2. Results.

Supplementary Figure 1. Serum cholesterol levels.

Supplementary Table 1. Overview of results from histopathological analysis of group 1 (n=9 rabbits) after 91 days.

Supplementary Table 2. Overview of results from OCT analysis of group 1 (n=9 rabbits) after 91 days.

Supplementary Table 3. Additional results from histopathological analysis of groups 2 and 3 (n=10 rabbits each) after 161 days.

Supplementary Table 4. Additional results from OCT analysis of groups 2 and 3 (n=10 rabbits each) after 161 days.

The supplementary data are published online at: https://eurointervention.pcronline.com/ doi/10.4244/EIJ-D-19-00747



Supplementary data

Supplementary Appendix 1. Methods

Study flow

Animals assigned to histology and optical coherence tomography

In a total of 33 rabbits, one BRS (n=33) and one permanent metallic DES (n=33) of equivalent geometry and design were randomly allocated to implantation into the right and left iliac arteries, respectively. Twenty-two rabbits were randomised to statin treatment (n=11, group 2) or placebo (n=11, group 3), while another 11 rabbits were terminated after 91 days (n=11, group 1) for evaluation of neoatherosclerosis formation prior to receiving statin therapy. All animals were fed a high-cholesterol diet (1% for 35 days followed by 0.025% until day 161). After seven days, endothelial balloon injury of the iliac arteries was performed using a Fogarty catheter, as previously described [21], followed by implantation of BRS and DES in accordance with random assignment. After 63 days, repeat balloon injury of the stented segment was performed to accelerate neoatherosclerosis formation. Animals of group 1 were terminated after 91 days while animals of groups 2 and 3 (n=11 each) were started on high-dose statin therapy (atorvastatin 3 mg/kg/d) or placebo and terminated after 161 days (**Figure 1**).

Animals assigned to scanning electron microscopy

Endothelialisation was investigated by scanning electron microscopy (SEM) in 12 rabbits (12 BRS and 12 DES) 28 days following stent implantation (day 35). Animals were fed an hypercholesterolaemic diet (1% cholesterol) throughout the entire in-life phase (**Figure 1**).

Study devices

In this study, a fully bioresorbable magnesium scaffold (Magmaris[®]; Biotronik AG, Bülach, Switzerland) was compared with a custom-designed 316L stainless steel drug-eluting stent (Biotronik AG) of equivalent geometry and design. Both BRS and DES were of the same size (3.0x15 mm), had a 6 crown 2 link design with a strut thickness of 150 μ m and used PLLA coating to elute sirolimus at a concentration of 140 μ g/cm².

Animal model of neoatherosclerosis

The rabbit model was approved by the government ("Regierung von Oberbayern", file number 55.2.1.54-2532-40-16) and was in accordance with the German Animal Welfare Act (version May 18, 2006, amended on March 29, 2017) as well as directive 2010/63/EU of the European Parliament on the protection of animals used for scientific purposes. Forty-five (45) New Zealand White rabbits (3.0 to 4.0 kg, 3 to 4 months of age; Charles River, France) were used for this preclinical animal model of in-stent neoatherosclerosis which has previously been established at our institution [13]. Animals were fed a 1% cholesterol diet (Altromin Spezialfutter GmbH, Lage, Germany) for seven days prior to balloon denudation of the iliac arteries, followed by stent implantation. After another four weeks of a high-cholesterol diet (1%), animals were switched to a 0.025% cholesterol diet (Altromin Spezialfutter GmbH) at day 35 and were continued on this diet until euthanasia. Blood samples were drawn on day 0, 7, 35, 63, 91, 126 and 161 for measurement of serum cholesterol and liver enzymes. Routine health checks including documentation of animal weight were performed three times a week.

Stent implantation, drug treatment, tissue harvest

On day 7, balloon denudation of the iliac arteries was performed using a 3 Fr Fogarty catheter before stents (3.0x15 mm; Biotronik AG) were implanted under fluoroscopic guidance at a nominal pressure of 10 atm (Magmaris) or 12 atm (316L SS-DES; Biotronik AG). Following implantation, vessel patency was documented by angiography. Animals received acetylsalicylic acid daily (40 mg/d) until the end of the experiment. Additionally, heparin (150 IU/kg) was administered during the procedure. On day 63, repeat denudation was performed to accelerate neoatherosclerosis formation. Animals assigned to SEM did not undergo repeat denudation procedures. Animals received buprenorphine (0.01 mg/kg) for a minimum of 24 hours after the interventions for post-procedural analgesia. On day 91, a subset of animals (n=11) was terminated to determine the magnitude of neoatherosclerosis formation prior to randomisation to statin treatment or placebo. The remaining animals (n=22) were randomised to statin (Atorvastatin Hennig[®] 10 mg) treatment (n=11) at an established dose of 3 mg/kg/d 10 per os or placebo (tap water per os) (n=11) and housed until day 161 to investigate changes in neoatherosclerosis progression over time. Euthanasia was induced by an overdose of pentobarbital, while rabbits were in deep anaesthesia. OCT imaging was performed immediately after euthanasia. Afterwards, iliac arteries were perfusion-fixed before they were processed for histopathological and immunohistochemical staining.

Histopathological and immunohistochemical processing

Stented arteries were embedded in methyl methacrylate (MMA). A total of five in-stent sections (10 µm thickness) were cut from each artery using a laser microtome (TissueSurgeon; LLS ROWIAK LaserLabSolutions GmbH, Hannover, Germany) and subsequently stained with haematoxylin-eosin (H&E) and Verhoeff-van Giesson (VVG). For immunohistochemistry, sections were stained using rabbit-specific antibodies against RAM-11 (Dako, Glostrup, Denmark, dilution 1:100) and SMA (Sigma Aldrich, St. Louis, MO, USA, dilution 1:150). Dako EnVision[™] Mouse Secondary Antibody kit was used. Staining was performed using a fully automated stainer (Leica Bond RXm; Leica Biosystems Nussloch GmbH, Nussloch, Germany).

Scanning electron microscopy

Twenty-eight days following stent implantation (day 35), 12 animals were terminated for evaluation of endothelial cell coverage by SEM. Tissue was harvested as described above. The samples were then dehydrated in a graded series of ethanol, critical point dried, and sputter-coated with gold. Vessels were visualised using a scanning electron microscope (EVO MA 15; Carl Zeiss, Oberkochen, Germany). Endothelial cells were identified as sheets of spindle or polygon-shaped monolayers in close apposition, a distinguishing feature in contrast to other cell types in en face preparations. Quantification of endothelial coverage was achieved with the help of a customised software algorithm (ImageJ 1.5; NIH, Bethesda, MD, USA). Strut endothelialisation was derived from the total area of endothelialisation minus the area between stent struts.

Histopathological assessment

Morphometric measurements were performed in VVG-stained slides, as previously described [22]. In H&E-stained slides, giant cell infiltration, haemorrhage and mineralisation were evaluated and expressed as percentage, i.e., (% giant cells = number of struts with giant cells/number of struts in cross-section x100). Furthermore, injury of the vessel wall was scored according to the Schwartz method [23] and inflammation was analysed according to the scoring system established by Otsuka et al [24]. Final average values per vessel were derived from all five in-stent sections.

Neoatherosclerosis score in histology

Infiltration of foamy macrophages was assessed in histologic sections using a dedicated scoring system. Infiltration of foamy macrophages was assessed in histologic sections after H&E staining using a quadrant-based scoring system comprising the depth of foam cell infiltration into the neointimal tissue along the vascular circumference - score 1: no foamy macrophage infiltration; score 2: foamy macrophage infiltration in superficial tissue layers; score 3: foamy macrophage infiltration in deeper tissue layers; score 4: foamy macrophages in both superficial and deeper tissue layers. Mean values per cross-section were derived from individual data points at each quadrant. Final average score per vessel was derived from all five in-stent sections.

OCT imaging, neoatherosclerosis scoring and morphometric analysis

Optical coherence tomography (OCT) was performed immediately after euthanasia using the frequency-domain C7-XR[™] OCT imaging console (OCT Intravascular Imaging System; St. Jude Medical, St. Paul, MN, USA). Image analysis was performed as previously described [22] using offline analysis (QIvus[®] Research Edition 3.0.26.0; Medis medical imaging systems bv, Leiden, the Netherlands). Analysis of OCT pullbacks included morphometric assessment of vessel dimensions and assessment of macrophage infiltration using an ordinal score.

For neoatherosclerosis scoring, OCT frames were evaluated for the presence of infiltrating macrophages with significant light attenuation [25] as well as hypo-intense areas with adjoining light attenuation. In every frame, the extent of infiltrating macrophages into the neointimal tissue was scored semi-quantitatively relative to the circumference of the vessel using an ordinal score - score 1: no macrophage infiltration; score 2: macrophage infiltration in <25% of the circumference; score 3: macrophage infiltration in 25-50% of the circumference; score 5: macrophage infiltration in >75% of the circumference. Final average score per vessel was derived from all in-stent OCT frames.

Morphometric measurements included lumen area, stent area and neointimal thickness over

struts. Neointimal area (stent area - lumen area) as well as percentage stenosis ([(stent area - lumen area)/stent area] x100) were calculated. Final average values per vessel were derived from every fifth in-stent OCT frame.

Statistical analysis

Continuous data are reported as mean with standard deviation in case of normal distribution and as median with interquartile range in case of non-parametric distribution. Distribution of data was confirmed by the Shapiro-Wilk goodness-of-fit test. The Wilcoxon Kruskal-Wallis rank-sum test was used to calculate the significance of differences between medians of nonparametric data, while ANOVA was used for group comparison of parametric data. To account for the clustered nature of data and the interaction of treatment assignments (BRS and DES assigned to statin and placebo), generalised estimating equation (GEE) modelling was used. For parametric data a linear model and for non-parametric data a gamma regression model with log link function was applied. Results are presented as estimated means with lower and upper 95% confidence interval (CI). To assess effects among local (BRS/DES) and systemic (statin/placebo) treatment, a p for interaction was calculated. To account for the categorical nature of the neoatherosclerosis score, Pearson's chi-square was also calculated for the quadrant-level comparison among treatment groups. Variability of scoring systems was calculated with intraobserver and interobserver correlation coefficients (ICCs) for absolute agreement. A p-value of p<0.05 was considered statistically significant. All analyses were carried out using JMP software, version 12.0 (SAS Institute, Cary, NC, USA) and SPSS, Version 22.0 (IBM Corp., Armonk, NY, USA).

Supplementary Appendix 2. Results

Results of group 1

Histopathological assessment of group 1 showed advanced neointimal coverage of stent struts in BRS relative to DES (% uncovered struts: 0.77% vs 33.64%, p=0.02). Neointima was thicker, but decreased foam cell infiltration was observed in BRS compared to DES (neointimal area: 3.01 mm² vs 1.89 mm², p=0.003; neoatherosclerosis score: 1.18 vs 1.43, p=0.0188) (**Supplementary Table 1, Supplementary Table 2**). Supplemental Figure 1



Supplementary Figure 1. Serum cholesterol levels.

Supplementary Table 1. Overview of results from histopathological analysis of group 1 (n=9 rabbits) after 91 days.

		A	Area EEL [m	m²]	Area IEL [mm²]			
		Mean	Std deviation	<i>p</i> -value	Mean	Std deviation	<i>p</i> -value	
Treatment	BRS, n=9	5.58	0.58	0.0054*	5.26	0.56	0.0013*	
Treatment	DES, n=9	6.44	0.55	0.0054	6.26	0.53	0.0015	
		Neo	intimal area	[mm ²]	Lumen area [mm ²]			
		Mean	Std deviation	<i>p</i> -value	Mean	Std deviation	<i>p</i> -value	
Treatment	BRS, n=9	3.01	0.59	0.0030*	2.24	0.72	0.0003*	
DES, n=9		1.89	0.76	0.0050	4.37	0.70	0.0005	
		N	ledia area [m	um²]	Injury score			
		Mean	Std deviation	<i>p</i> -value	Mean	Std deviation	<i>p</i> -value	
Treatment	BRS, n=9	0.32	0.08	0.0005*	0.41	0.27	0 5660	
Treatment	DES, n=9	0.18	0.04	0.0005	0.48	0.27	0.5000	
		Neoatherosclerosis score			Inflammation score			
		Mean	Std deviation	<i>p</i> -value	Mean	Std deviation	<i>p</i> -value	
Treatment	BRS, n=9	1.18	0.08	0.0188*	1.84	0.76	0 1092	
Troumont	DES, n=9	1.43	0.22	0.0100	1.31	0.49	0.1092	
		% Stenosis			% Uncovered struts			
		Mean	Std deviation	<i>p</i> -value	Mean	Std deviation	<i>p</i> -value	
Treatment	BRS, n=9	58.21	11.25	0.0007*	0.77	1.59	0.0200*	
reatment	DES, n=9	30.07	10.63	0.0007	33.64	27.88	0.0200*	

Values are presented as means with standard deviation. A p<0.05 was considered statistically significant (*).

Supplementary Table 2. Overview of results from OCT analysis of group 1 (n=9 rabbits) after 91 days.

		\mathbf{L}	umen area [n	nm²]	Stent area [mm ²]			
		Mean	Std deviation	<i>p</i> -value	Mean	Std deviation	<i>p</i> -value	
Treatment	BRS, n=9	BRS, n=9 2.54 0.51 0.0013		0.0013*	4.51	0.48	0.0017*	
Treatment	DES, n=9	3.80	0.50	0.0015	5.69	0.51	0.0017	
		Neo	intimal area	[mm ²]	Neointimal thickness over struts			
		Mean	Std deviation	<i>p</i> -value	Mean	Std deviation	<i>p</i> -value	
Traatmont	BRS, n=9	1.86	0.58	0.0103*	0.33	0.10	0.00/1*	
Treatment	DES, n=9	1.09	0.64	0.0195	0.16	0.08	0.0041	
			% Stenosis	3	Neoatherosclerosis score			
		Mean	Std deviation	<i>p</i> -value	Mean	Std deviation	<i>p</i> -value	
Treatment	BRS, n=9	43.27	14.10	0.0031*	1.13	0.18	0.123	
Treatment	DES, n=9	18.94	10.81	0.0031	1.32	0.32	0.125	

Values are presented as means with standard deviation. A p<0.05 was considered statistically significant (*).

Area EEL [mm²] Area IEL [mm²] Lower Upper Lower Upper B-Mean 95% 95% *p*-value **B**-value Mean 95% 95% *p*-value value CI CI CI CI BRS. 4.92 4.69 5.16 4.69 4.46 4.93 n=10 Treat 0.0001* -0.282 0.0001* -0.307 DES, ment 6.55 6.42 6.68 6.37 6.24 6.50 n=10 Statin, 5.63 5.38 5.89 5.42 5.17 5.69 n=10 -0.011 -0.016 0.616 0.624 Group Placebo, 5.72 5.49 5.96 5.51 5.27 5.75 n=10 p for interaction 0.828 0.994 Neointimal area [mm²] Lumen area [mm²] Lower Upper Lower Upper B*p*-value Mean 95% 95% **B**-value Mean 95% 95% *p*-value value CI CI CI CI BRS, 3.22 2.96 3.48 1.47 1.28 1.69 n=10 Treat 0.0001* 0.987 0.0001* -1.041 ment DES, 2.27 2.04 2.50 4.10 3.91 4.30 n=10 Statin, 2.72 2.34 3.10 2.44 2.20 2.71 n=10 0.851 -0.005 0.912 -0.024 Group Placebo, 2.77 2.51 3.03 2.46 2.17 2.80 n=10 p for interaction 0.662 0.817 Media area [mm²] % Uncovered struts Lower Upper Lower Upper B-Mean 95% 95% *p*-value **B**-value Mean 95% 95% *p*-value value CI CI CI CI BRS, 0.23 0.21 0.25 0.00 0.00 0.00 Treat n=10 0.0001* 0.078 0.0001* -12.03 DES, ment 0.18 0.16 0.20 15.58 8.98 27.02 n=10 Statin, 0.21 0.19 0.23 0.038 0.025 0.058 n=10 0.807 0.031 0.786 -0.153 Group Placebo, 0.20 0.18 0.23 0.041 0.029 0.059 n=10 p for interaction 0.044 0.786 % Giant cells Inflammation score Lower Lower Upper Upper B-Mean 95% 95% *p*-value **B**-value Mean 95% 95% *p*-value value CI CI CI CI BRS, 1.53 1.31 1.75 9.33 6.76 11.90 n=10 Treat 0.106 -0.057 0.335 0.574 ment DES, 1.24 1.01 1.47 7.70 5.92 9.47 n=10 Statin, 1.27 1.65 6.65 11.15 1.46 8.90 n=10 0.293 -0.200 0.601 -0.284 Group Placebo, 1.31 1.11 1.51 8.13 6.29 9.96 n=10 p for interaction 0.054 0.786 **Injury score** Upper Lower Mean 95% **B**-value

95%

CI

CI

p-value

Supplementary Table 3. Additional results from histopathological analysis of groups 2 and 3 (n=10 rabbits each) after 161 days.

Treat	BRS, n=10	0.40	0.35	0.45	0.776	0.046			
ment	DES, n=10	0.42	0.34	0.49	0.776	-0.040			
Group	Statin, n=10	0.43	0.36	0.51	0.311	0.014			
Gloup	Placebo, n=10	0.39	0.34	0.44	0.311	0.014			
<i>p</i> for interaction 0.426									

Values are presented as estimated means with upper and lower 95% CI. A p<0.05 was considered statistically significant (*).

Lumen area [mm ²]							Stent area [mm ²]				
		Mean	Lower 95% CI	Upper 95% CI	<i>p</i> -value	B- value	Mean	Lower 95% CI	Upper 95% CI	<i>p</i> -value	B- value
Treat	BRS, n=10	2.08	1.81	2.39	0.0001*	-0.875	4.08	3.75	4.44	0.0001*	-0.412
ment	DES, n=10	4.61	4.41	4.82			5.97	5.82	6.12		
	Statin, n=10	3.16	2.89	3.45			4.99	4.70	5.29		
Group	Placebo, n=10	3.04	2.70	3.41	0.596	-0.038	4.88	4.52	5.28	0.660	-0.009
. for	•						0.422				
<i>p</i> 10r	interaction		0	.302				0	.422		
<i>p</i> 10r	Interaction		0 Neoin	.302 ntimal area	a [mm ²]		N	0 eointimal t	.422 hickness o	ver struts [m	m]
<i>p</i> 10r	Interaction	Mean	0 Neoin Lower 95% CI	atimal area Upper 95% CI	a [mm²] <i>p</i> -value	B- value	N Mean	eointimal t Lower 95% CI	422 hickness o Upper 95% CI	ver struts [m <i>p</i> -value	m] B- value
Treat	BRS, n=10	Mean 1.86	0 Neoin Lower 95% CI 1.63	302 ntimal area Upper 95% CI 2.09	a [mm ²]	B- value	N Mean 0.34	eointimal t Lower 95% CI 0.30	422 hickness o Upper 95% CI 0.39	ver struts [m <i>p</i> -value	m] B- value
Treat	BRS, n=10 DES, n=10	Mean 1.86 1.32	0 Neoin 25% CI 1.63 1.12	302 ntimal area 95% CI 2.09 1.52	a [mm ²] <i>p</i> -value 0.0001*	B- value 0.611	N Mean 0.34 0.19	0 eointimal (95% CI 0.30 0.16	422 hickness o Upper 95% CI 0.39 0.22	ver struts [m <i>p</i> -value 0.0001*	m] B- value 0.666
Treat	BRS, n=10 DES, n=10 Statin, n=10	Mean 1.86 1.32 1.62	0 Neoin 25% CI 1.63 1.12 1.33	302 ntimal area 95% CI 2.09 1.52 1.92	a [mm ²] <i>p</i> -value 0.0001*	B- value 0.611	N Mean 0.34 0.19 0.26	0 eointimal t 95% CI 0.30 0.16 0.22	422 hickness o Upper 95% CI 0.39 0.22 0.31	ver struts [m <i>p</i> -value 0.0001*	m] B- value 0.666
Treat ment Group	BRS, n=10 DES, n=10 Statin, n=10 Placebo, n=10	Mean 1.86 1.32 1.62 1.56	0 Neoin 95% CI 1.63 1.12 1.33 1.38	.302 ntimal area Upper 95% CI 2.09 1.52 1.92 1.74	a [mm ²] <i>p</i> -value 0.0001* 0.723	B- value 0.611 0.133	N Mean 0.34 0.19 0.26 0.25	0 eointimal t 95% CI 0.30 0.16 0.22 0.22	422 hickness o Upper 95% CI 0.39 0.22 0.31 0.28	ver struts [m <i>p</i> -value 0.0001* 0.639	m] B- value 0.666 0.133

Supplementary Table 4. Additional results from OCT analysis of groups 2 and 3 (n=10 rabbits each) after 161 days.

Values are presented as estimated means with upper and lower 95% CI. A p<0.05 was considered statistically significant (*).