EuroIntervention

Porcine models of coronary atherosclerosis and vulnerable plaque for imaging and interventional research

Juan F. Granada^{1*}, MD, FACC; Greg L. Kaluza¹, MD, PhD, FACC; Robert L. Wilensky², MD; Barbara C. Biedermann³, MD; Robert S. Schwartz⁴, MD, FACC; Erling Falk⁵, MD, PhD

 Skirball Center for Cardiovascular Research, Cardiovascular Research Foundation, Orangeburg, NY, USA; 2. Hospital of the University of Pennsylvania, Philadelphia, PA, USA; 3. Department of Medicine, University Hospital Bruderholz, Bruderholz, Switzerland; 4. The Minneapolis Heart Institute, Abbott Northwestern Hospital, Minneapolis, MN, USA;
Atherosclerosis Research Unit, Aarhus University Hospital, Aarhus, Denmark

The authors have no conflict of interest to declare.

KEYWORDS

Angiogenesis, coronary artery disease, plaque rupture, complex lesions, imaging

Abstract

Animal models facilitate our understanding of human disease by providing a controlled environment permitting testing of mechanisms of disease, diagnostic technologies and therapeutic interventions. The ideal animal model should display coronary lesions resembling those seen in human atherosclerosis. No suitable large animal model of high-risk (vulnerable) plaque exists. Lack of such a model has hampered studies designed to validate imaging technologies and to scrutinise the effects of therapeutic interventions in atherosclerotic arteries. Several porcine models of advanced human-like coronary atherosclerosis exist. In this review some of the most promising porcine models are discussed, focusing on their applicability in the development and validation of coronary imaging technologies and interventional devices. In the evolving era of technological development, the availability and use of such animal models of advanced human-like coronary atherosclerosis and vulnerable plaque will become critically important in the preclinical testing of emerging technologies in interventional cardiology.

* Corresponding author: Skirball Center for Cardiovascular Research, Cardiovascular Research Foundation, 8 Corporate Drive, Orangeburg, NY, 10962, Orangeburg, NY, USA
E-mail: jgranada@crf.org
© Europa Edition. All rights reserved.

PCRONI INF COM

EuroIntervention 2009;5:140-148 published online ahead of print March 2009



Introduction

Animal models facilitate the understanding of human disease when important aspects of the disease process develop in a predictable time period within a controlled environment. Although no animal model can fully replicate complex human pathological conditions, animal models are key for the evaluation of mechanisms of disease and testing of diagnostic technologies and therapeutic interventions¹⁻³. Specifically, animal models of atherosclerosis and high-risk (vulnerable) plaque are pivotal in the development and validation of novel imaging technologies. Such validation has been hampered by the lack of a suitable large animal model of humanlike coronary atherosclerosis^{1,4}. Historically, non-atherosclerotic porcine coronary injury models have been used extensively in the validation of treatments against restenosis, and device development. However, current diagnostic and therapeutic devices should be tested in more clinically relevant animal models. Pigs harbouring metabolic conditions such as hypercholesterolaemia or diabetes develop atherosclerotic plaques in anatomical locations that are similar to the human condition. Depending on swine breed, cholesterol levels, exposure time or the presence of vascular injury, advanced and heterogeneous coronary lesions displaying humanlike features may develop in a relatively short period of time. This paper reviews the current status of several porcine models of coronary atherosclerosis and vulnerable plaque and discusses the applicability of these models for testing and validation of emerging coronary imaging technologies.

Swine models in biomedical research

Genetically, the pig is relatively close to humans⁵. This also applies to the anatomy and physiology of the cardiovascular system and the arterial response to hypercholesterolaemia. The domestic crossbred farm pig, sus scrofa domestica, is the most commonly used swine in cardiovascular research today^{2,3,6}. However, in situations in which significant growth would be problematic, miniature swine (Yucatan, Hanford, Gottingen and Sinclair Hormel breeds) are often used in chronic studies. Typically, swine achieve sexual maturity by six to eight months at which time their weight usually varies between 40 and 100 kg. The heart of the pig is anatomically similar to the human heart with the exception of having a left azygous vein draining the intercostal system into the coronary sinus⁷ and a right coronary artery ostium situated substantially more cranially. The heart of a 40-50 kg miniature pig is approximately the same size as an adult human heart⁸. The coronary artery system is similar to 90% of the

human population in anatomy and function, is more prone to vasospasm during manipulation, and possesses no pre-existing collaterals between the coronary arteries and their branches⁹.

Developmental aspects of swine models for atherosclerosis

Several important biological and anatomical factors must be considered before selecting the appropriate animal model in the setting of technology validation. For testing of emergent imaging techniques, the developed atherosclerotic lesions should contain the basic histological components of interest, such as a large necrotic core, a thin fibrous cap, inflammatory cells, and/or angiogenesis^{10,11}. Alternatively, for stent validation studies, more advanced and fibrotic coronary lesions may be more desirable. Focal atherosclerotic lesions located in accessible regions are desirable. The reproducibility of the model should avoid the screening of multiple animals in an attempt to isolate a cohort sharing desired anatomical features. Normal swine on normal feed have low plasma LDL and relatively high HDL cholesterol levels and, consequently, do not develop mature atherosclerotic lesions. However, when fed a cholesterol-rich diet swine will develop hypercholesterolaemia (±300 mg/dl) and atherosclerotic lesions similar to those seen in humans. The evolution and histological types of lesion developed depend on swine breed, cholesterol level, exposure time, and coexisting proatherogenic conditions. Complex atheromatous lesions may eventually develop, but usually require long development times including months of severe hypercholesterolaemia. Although atherosclerotic plaque rupture and luminal thrombosis occur they are uncommon¹²⁻¹⁴. As in humans, diet-induced atherosclerosis in swine is multifactorial and results in the development of multifocal lesions of variable composition in unpredictable locations. A significant challenge facing animal model development is to shorten the typical 6-9 month period required to develop complex atherosclerotic lesions. Through the appropriate selection of swine breed, animals highly sensitive to dietary manipulation have been shown to develop complex lesions in a slightly accelerated time period (Table 1).

Porcine inherited hyper-LDL-cholesterolaemia model

Rapacz et al originally described a strain of large domestic swine which develops elevated LDL cholesterol levels and spontaneous atherosclerosis while on a low-fat diet due to a mutation in genes

| Table 1. Key developmental aspects of porcine atherosclerotic r |
|---|
|---|

| | HCD | FHS without HCD | Diabetes + HCD | Injury + HCD |
|---------------------|-----------------------|-------------------------------|--------------------------|----------------------|
| Anatomical location | Coronary, aorta, | Coronary, aorta, | Coronary, aorta, | Injury-dependent |
| | iliofemoral | iliofemoral | iliofemoral | Localised lesion(s) |
| Advanced lesions | >2 years | 12-18 months | 6-9 months | 9-12 months |
| Advantages | Easy | Endogenous HS | Accelated model | Predictable location |
| | Universally available | No costly diet | Less weight gain | Accelerated model |
| Major limitations | Long-term model | Long-term model | Failure to thrive | Complicated |
| | Costly diet | Limited availability | Costly (STZ, healthcare) | Costly procedures |
| | Unmanagable size* | Unmanagable size [#] | Not short-term model | Not short-term model |

HCD: High cholesterol diet; FHS: Familiar hypercholesterolemic swine; STZ: streptozotocin; * Does not apply to minipigs; # Does not apply to down-sized FHS pigs



coding for apolipoproteins and/or the LDL receptor^{15,16}. As a result these pigs develop human-like atherosclerotic lesions on normal feed. By 12 months of age, early atherosclerotic lesions composed of macrophage-derived foam cells and smooth muscle cells are seen in the most susceptible arteries (coronary and iliofemoral arteries and aorta). However, a rapid progression of disease between 12 and 18 months is observed and well-developed atheromas are found in 60% of the pigs by 18 months (Figure 1). Peripheral arterial lesions are more frequently found in the thoracic aorta and aorto-iliac bifurcation and tend to be more fibrous than the coronary lesions. By 24 months, complicated stenotic lesions containing fibrous caps, necrotic cores, cholesterol clefts, granular calcium deposits, and neovascularisation deep within the lesion are almost universally present in the major coronary arteries (Figure 2). By 36 months, plaque neovascularisation and haemorrhage are common, and plaque rupture may occur. Luminal thrombosis appears to be rare and acute occlusive thrombosis has not been documented, however, sudden cardiac death is common beyond three years of follow-up (personal communication JFG).

Atherosclerotic lesions in this model are characterised by marked plaque neovascularisation, a finding that can be exploited to test imaging modalities that detect this feature. Necrotic core formation and eccentricity of lesions in this model are also very similar to human disease, however, media destruction is comparatively less pronounced. Although this swine model develops humanlike atherosclerosis while maintained on a regular diet, a major limitation is the duration necessary for advanced coronary lesions to develop (>1 year) and the subsequent difficulties encountered in handling the mature animals, whose weights can exceed 200 kg. A cholesterol enriched diet might accelerate the process and thus mitigate these limitations but, unfortunately, no such information is available. Ongoing efforts are being undertaken both in the US and Europe to downsize the spontaneously hypercholesterolaemic pig to a more manageable size which will increase its usefulness substantially.

Porcine diabetes/hypercholesterolaemia model

Gerrity et al demonstrated that the combination of induced diabetes and hypercholesterolaemia (DM/HC) in Yorkshire pigs led to advanced atherosclerotic lesions in the coronary, femoral arteries and aorta in a relatively short period of time (20 weeks)¹⁷. Diabetes was induced with an intravenous infusion of streptozotocin resulting



Figure 1. Natural history of atherosclerosis in the familiar hypercholesterolaemic swine (inherited hyper-LDL-cholesterolaemia). Lipid-rich lesions are stained by Oil Red O. A: Time-dependent development of lesions in the left main, left anterior descending (LAD) coronary artery and circumflex branch. At 16 months of age, advanced lesions are predominantly seen proximal in the coronary arteries. B: Disease distribution: At 18 months, lesion development is predominantly in the proximal thoracic aorta, distal abdominal aorta and its trifurcation, and proximal coronary arteries (arrows). (Figure courtesy of Chris Krueger and Jess Reed at the Department of Animal Sciences, University of Wisconsin, Madison).





Figure 2. Advanced human-like coronary lesions in familial hypercholesterolaemic swine (inherited hyper-LDL-cholesterolaemia). Left: Atheromatous lesion containing a large necrotic core (NC) covered by a fibrous cap (FC) that is very thin near the shoulder region (vulnerable plaque). Right: Complex stenotic lesion containing collagen, lipid, calcification, and inflammation, predominantly macrophage foam cells. Collagen is blue in left panel (Masson stain), green in right panel (Movat stain).

in a >80% reduction in pancreatic beta cells. Following administration of a high cholesterol diet, accelerated atherosclerosis was observed. Diabetic only animals did not develop atherosclerosis¹⁷ but the combination of diabetes and hypercholesterolaemia led to accelerated development of more advanced lesions. Early atherosclerotic lesions developing within three months are generally intimal xanthomas¹⁸. By six months, 96% of arteries harbour an atherosclerotic lesion with progression to 100% at nine months post induction. At 9-months many arteries demonstrated complex human-like lesions (11%, Figure 3)^{17,18}. As in humans, the most severe lesions were generally located in the proximal segment of the coronary arteries often at the side-branch

points. Chatzizisis et al demonstrated that areas of low endothelial shear stress in coronary arteries were associated with subsequent development of lesions. These lesions had increased lipid accumulation, inflammation and expansive remodelling; all characteristics of high-risk (vulnerable) atherosclerotic lesions¹⁹. Focal calcifications were noted at 6-9 months as well as thin-cap fibroatheromas in some arteries. More advanced lesions possessed acellular necrotic cores covered by fibrous caps with medial thinning, haemorrhage and calcification (11% thin-cap fibroatheromas)¹⁸. Treatment with darapladib, a selective inhibitor of Lp-PLA2, an important mediator of vascular inflammation has been shown to reduce development of atherosclerosis but more importantly necrotic core development in this model²⁰.



Figure 3. Advanced lesions in diabetic+hypercholesterolaemic (DM/HC) swine. Left, 6 months after DM/HC induction: Complex stenotic plaque in an iliac artery containing well-developed necrotic cores (NC) covered by a fibrous cap (FC). Intraplaque haemorrhage (H) is also seen. Arrow indicates a side-branch and tunica media (M). Right, 9 months after DM/HC induction: Coronary fibroatheroma with large necrotic cores (NC) and heavy calcifications near the internal elastic lamina which is destroyed from 8 to 10 o'clock and at 12 o'clock. The lumen (*) is severely narrowed. Both Movat stain.



As with humans the development of atherosclerosis in DM/HC is variable with regard to location, severity and extent. Coronary arteries develop lesions more often and more severe than within the peripheral arterial territory^{17,18}. Carotid arteries showed more resistance to lesion development under DM/HC conditions. The proximal iliac artery is also predisposed to lesion development (Figure 3). Studies of gene expression have shown that inflammatory genes were more markedly upregulated in coronary arteries than thoracic aorta and carotid arteries from the same animal¹⁸. This may reflect the presence of pulsatile flow and significant number of branching arteries in the coronary circulation. A distinct advantage to the DM/HC model is that diabetic pigs gain weight at a slower rate than hyperlipidaemic or non-diseased pigs. Limitations of the model are the variability in size and location of the developed lesions and the expense in maintaining the model. In addition, DM/HC animals can develop hypoglycaemic coma if not closely monitored and are prone, as are human diabetics, to infections and gastroparesis. A DM/HC model has also been described in Yucatan minipigs, but plaque size was only reported for a few $pigs^{21}$.

Minipig models of coronary atherosclerosis

Atherosclerosis is a chronic disease, and it takes relative long time to develop advanced lesion in pigs similar to those responsible for clinical disease in humans. The rapid growth of the domestic swine makes it less suitable for long-term studies compared to minipigs, not only because of the high maintenance costs but also because of the continued growth (non-steady coronary dimensions) leading to a large and heavy pig that is difficult to image (loss of resolution) and handle. Some colonies of Yucatan minipigs have proved to be susceptible to both diet-induced hypercholesterolaemia and atherosclerosis, and they have been used for decades in atherosclerosis and restenosis research^{13,22-25}. Complicated human-like atherosclerotic lesions displaying necrosis, cholesterol crystals, and calcification may develop relatively rapidly on a cholesterol-rich atherogenic diet¹³. Furthermore, streptozotocininduced diabetes has been described, and preliminary data indicate that it accelerates diet-induced coronary atherosclerosis and the development of vulnerable plaques in Yucatan minipigs as it does in Yorkshire swine²¹. Other minipigs, such as the Göttingen, Hanford and Sinclair Hormel breeds, have also been used in atherosclerosis research but far less information concerning their use is available compared to the Yucatan mini/micro swine. Despite great enthusiasm a few years ago, human-like atherosclerosis has not yet been documented in the obese, insulin-resistant, mildly hypertensive, hypercholesterolaemic Ossabaw pig²⁶.

Porcine vascular injury/hypercholesterolaemia models

In order to shorten the time required to develop lesions of interest and limit the high maintenance costs, multiple attempts have been made to accelerate the atherosclerotic process by combining dietinduced hypercholesterolaemia with a localised vascular injury such as external radiation²⁷, wire-induced endothelial denudation²⁸, barotrauma^{24,25,29}, or haemodynamic manipulations³⁰. In 2001,

Keelan and Schwartz introduced the concept of "accelerating" the atherosclerotic process locally by combining balloon injury (Infiltrator catheter, Boston Scientific, Minneapolis, MN, USA) and triggering inflammation by directly injecting a large lipid pool into the vessel wall of hypercholesterolaemic pigs³¹. One variation of this approach involves the delivery of a large concentration of lipids into the adventitial layer of normal porcine coronary arteries using a specialized micro-needle catheter³². With this technique excessive vascular trauma secondary to overstretching was avoided by direct injection. Feasibility studies demonstrated safety and efficacy of this technique³². Short term studies (eight weeks) demonstrated that intramural delivery of lipids initiate the process of atherosclerotic disease. By seven weeks, IVUS analysis of the injected segment showed that the lipid injection induces the formation of early eccentric fibrotic lesions located in positively remodelled segments. By histology, the lesions are eccentric, nonobstructive and contain large amounts of lipid co-localising with foam cells. After 12 weeks of delivery, the lesions continue to develop and are more significant in size and contain increased amounts of lipid and macrophages³³. At 12 months, these atherosclerotic lesions show a necrotic core covered by a dense fibrous cap containing foam cells and abundant intraplaque neovessels. The locally induced lesions in this model are characterised by the rupture of the elastic components of the vessel and abundant adventitial plaque neovascularisation (Figure 4). Interestingly, due to the high cholesterol diet, staining of the surface of the vessels usually displays early atherosclerosis in the thoracic aorta, distal abdominal aorta and proximal coronary arteries. Compared to balloon injury or saline injection, the injected vascular segments demonstrate increased expression of chemokine mRNA and chemokine receptors important in macrophage recruitment³⁴. An important feature of this model is that the anatomical distribution of the lesions is predictable as it follows the pattern of injury induced. Therefore, it provides an opportunity for selecting the anatomical targets and optimising the sample size. The major limitation of this model is the complexity of lesion induction requiring mechanical intervention and diet supplementation. Although the development time is reduced, it still takes up to one year until advanced coronary lesions are evident. Additionally, the barotrauma associated with the delivery device may produce a disproportionate component of neointimal hyperplasia³⁵. In order to decrease the development time even further, several groups are attempting to accelerate the atherosclerotic process by inducing mechanical injury in the porcine inherited hyper-LDLcholesterolaemic model.

Research applications of swine models of atherosclerosis

In this review we have tried to highlight the fact that pig models may develop humanlike coronary atherosclerosis if exposed to atherogenic stimuli for at least several months. Plaque size and composition depend on the method used to induce the lesion. High-risk plaque features assumed to confer "vulnerability" in humans are also seen in advanced porcine lesions, including a large necrotic core, a thin fibrous cap, inflammation,





Figure 4. Advanced coronary lesions at predictable location (proximal coronary arteries) after local injury (lipid injection) in hypercholesterolaemic swine. As illustrated, coronary fibroatheromas with necrotic cores covered by fibrous caps are common in this model. Inflammatory cells, including macrophage foam cells, are usually seen in the fibrous cap at high magnification (not shown). Collagen is green-blue in this polychrome staining.

angiogenesis and plaque haemorrhage. The ultimate proof of "vulnerability" is, however, usually lacking, because plaque rupture is uncommon and luminal thrombosis is rare. Despite this deficiency, porcine models of coronary atherosclerosis and morphologically vulnerable plaques can provide meaningful information in the development, validation, and improvement of novel diagnostic and therapeutic technologies for use in interventional cardiology.

Novel imaging technologies to detect atherosclerosis, specifically high-risk (vulnerable) coronary artery lesions, are being evaluated in both humans and animal models. For validating non-invasive diagnostic technology, swine models provide suitable size, vascular anatomy and physiology similar to human disease. Indeed, the majority of the initial validation work on CT angiography and contrast media injection and MRA compatible contrast agents^{36,37} has been developed in the normal porcine model³⁸⁻⁴¹. In this field, porcine models have been previously used in studies requiring the presence of arterial obstructions^{42,43} or the development of acute myocardial infarction⁴⁴. Particularly, porcine models have been very useful in the evaluation of PET related technologies⁴⁵⁻⁴⁸ and molecular tracers⁴⁹⁻⁵¹. In the field of invasive cardiovascular imaging, animal models are relevant as it is important to document and characterise morphology, location and presence of specific structural plaque components such as calcium. Most of the initial IVUS⁵² and OCT technology development^{53,54} involved the use of the normal porcine coronary model. However, as the endovascular imaging technology field evolves, the use of the normal porcine coronary model will be no longer viable. Specifically, the characterisation of specific biological components within the vessel wall including remodeling⁵⁵, plaque strain⁵⁶ or tissue components⁵⁷ have required the use of injury based animal models. These models may be critical during the validation process of imaging technologies designed to detect specific biological components (i.e., IVUS based tissue characterisation and vasa vasorum imaging) encountered in human atherosclerotic lesions (Figure 5). Finally, in the area of cardiovascular interventions, healthy swine have been traditionally used as the animal model to test endovascular technology. In the evolving era of drug eluting stents, for example, a large animal with pre-existing metabolic and/or arterial disease may be needed to validate potential therapeutic strategies and their long-term consequences, such as demonstrating the effects of therapies directed toward limiting vascular inflammation and necrosis, preserving the endothelium, and promoting healing^{23,58,59}.

Conclusions

Clinical implementation of innovative diagnostic and therapeutic technologies will ultimately depend on the successful development of large animal models of humanlike coronary atherosclerosis that permit preclinical validation of the technology in a controlled environment. General limitations in porcine model development are well recognised and include the need for significant infrastructure for animal maintenance, the long time period needed to induce and monitor the lesions, and development costs. In current models of "naturally" occurring atherosclerosis (genetic, diet-induced), significant coronary artery disease occurs late and is unpredictable in location. Hence, further model development is needed, including innovative ways to accelerate the atherosclerotic process in susceptible animals with or without pre-existing arterial disease. The current limitations notwithstanding, valuable porcine models of atherosclerosis are already available for development, validation, and improvement of imaging technologies. How reliably these porcine models mimic the chronic pathobiology and natural history of human atherosclerosis requires further exploration. Regardless, the availability of animal models of human-like atherosclerosis will become perhaps the most critical element of the preclinical validation of emerging diagnostic and therapeutic technologies developed for use in patients with coronary artery disease.

Acknowledgements

The authors are thankful and extend gratitude for the research collaboration and support of Chris Krueger and Jess Reed at the Department of Animal Sciences, University of Wisconsin, Madison. The authors would like to thank David Wallace-Bradley for his assistance in the preparation and editing of this manuscript.





Figure 5. IVUS-based tissue characterisation performed in swine. A non-obstructive lesion is visualised in the proximal segment of the RCA located in a previously injured segment. After injury, swine were maintained on a high cholesterol diet for 9 months. Plaque evaluation by QCA (A), IVUS (B), IVUS-based tissue characterisation (C) and histology (D).

References

1. Rekhter MD. How to evaluate plaque vulnerability in animal models of atherosclerosis? *Cardiovasc Res.* 2002;54(1):36-41.

2. Schwartz RS, Edelman ER, Carter A, Chronos N, Rogers C, Robinson KA, Waksman R, Weinberger J, Wilensky RL, Jensen DN, Zuckerman BD, Virmani R. Drug-eluting stents in preclinical studies: recommended evaluation from a consensus group. *Circulation*. 2002;106(14):1867-1873.

3. Schwartz RS, Edelman ER, Carter A, Chronos NA, Rogers C, Robinson KA, Waksman R, Machan L, Weinberger J, Wilensky RL, Goode JL, Hottenstein OD, Zuckerman BD, Virmani R. Preclinical evaluation of drugeluting stents for peripheral applications: recommendations from an expert consensus group. *Circulation*. 2004;110(16):2498-2505.

4. Granada JF, Kaluza GL, Raizner AE, Moreno PR. Vulnerable plaque paradigm: prediction of future clinical events based on a morphological definition. *Catheter Cardiovasc Interv.* 2004;62(3):364-374.

5. Wernersson R, Schierup MH, Jorgensen FG, Gorodkin J, Panitz F, Staerfeldt HH, Christensen OF, Mailund T, Hornshoj H, Klein A, Wang J, Liu B, Hu S, Dong W, Li W, Wong GK, Yu J, Bendixen C, Fredholm M, Brunak S, Yang H, Bolund L. Pigs in sequence space: a 0.66X coverage pig genome survey based on shotgun sequencing. *BMC Genomics*. 2005;6(1):70.

6. Schook L, Beattie C, Beever J, Donovan S, Jamison R, Zuckermann F, Niemi S, Rothschild M, Rutherford M, Smith D. Swine in biomedical research: creating the building blocks of animal models. *Anim Biotechnol.* 2005;16(2):183-190.

7. Swindle MM, Horneffer PJ, Gardner TJ, Gott VL, Hall TS, Stuart RS, Baumgartner WA, Borkon AM, Galloway E, Reitz BA. Anatomic and anesthetic considerations in experimental cardiopulmonary surgery in swine. *Lab Anim Sci.* 1986;36(4):357-361.

8. Smith AC, Spinale FG, Swindle MM. Cardiac function and morphology of Hanford miniature swine and Yucatan miniature and micro swine. *Lab Anim Sci.* 1990;40(1):47-50.



9. White FC, Carroll SM, Magnet A, Bloor CM. Coronary collateral development in swine after coronary artery occlusion. *Circ Res.* 1992;71(6):1490-1500.

10. Fleiner M, Kummer M, Mirlacher M, Sauter G, Cathomas G, Krapf R, Biedermann BC. Arterial neovascularization and inflammation in vulnerable patients: early and late signs of symptomatic atherosclerosis. *Circulation*. 2004;110(18):2843-2850.

11. Falk E. Pathogenesis of atherosclerosis. *J Am Coll Cardiol.* 2006;47(8 Suppl):C7-12.

12. Lee KT, Kim DN, Thomas WA. Atherosclerosis in Swine. In: Stanton HC, Mersmann HJ, eds. Swine in Cardiovascular Research. Boca Raton, FL: CRC; 1986:33-48.

13. Reitman JS, Mahley RW, Fry DL. Yucatan miniature swine as a model for diet-induced atherosclerosis. *Atherosclerosis*. 1982;43(1):119-132.

14. Thorpe PE, Hunter WJ, 3rd, Zhan XX, Dovgan PS, Agrawal DK. A noninjury, diet-induced swine model of atherosclerosis for cardiovascular-interventional research. *Angiology.* 1996;47(9):849-857.

15. Hasler-Rapacz J, Ellegren H, Fridolfsson AK, Kirkpatrick B, Kirk S, Andersson L, Rapacz J. Identification of a mutation in the low density lipoprotein receptor gene associated with recessive familial hypercholesterolemia in swine. *Am J Med Genet.* 1998;76(5):379-386.

16. Prescott MF, McBride CH, Hasler-Rapacz J, Von Linden J, Rapacz J. Development of complex atherosclerotic lesions in pigs with inherited hyper-LDL cholesterolemia bearing mutant alleles for apolipoprotein B. *Am J Pathol.* 1991;139(1):139-147.

17. Gerrity RG, Natarajan R, Nadler JL, Kimsey T. Diabetes-induced accelerated atherosclerosis in swine. *Diabetes*. 2001;50(7):1654-1665.

18. Mohler ER, 3rd, Sarov-Blat L, Shi Y, Hamamdzic D, Zalewski A, Macphee C, Llano R, Pelchovitz D, Mainigi SK, Osman H, Hallman T, Steplewski K, Gertz Z, Lu MM, Wilensky RL. Site-specific atherogenic gene expression correlates with subsequent variable lesion development in coronary and peripheral vasculature. *Arterioscler Thromb Vasc Biol.* 2008;28(5):850-855.

19. Chatzizisis YS, Coskun AU, Jonas M, Edelman ER, Feldman CL, Stone PH. Role of endothelial shear stress in the natural history of coronary atherosclerosis and vascular remodeling: molecular, cellular, and vascular behavior. *J Am Coll Cardiol.* 2007;49(25):2379-2393.

20. Wilensky RL, Shi Y, Mohler ER, III, Hamamdzic D, Burgert ME, Li JU, Postle A, Fenning RS, Bollinger JG, Hoffman BE,Pelcovitz D, Yang J, Webb C, Zhang L, Zhang P, Gel b MH, Walker MC, Zalewski A, Macphee CH. Inhibition of lipoprotein-associated phospholipase A2 reduces complex coronary atherosclerotic plaque development. *Nat Med* 2008; 14:1059-1068.

21. Wang YX, Fitch R, Li W, Werner M, Halks-Miller M, Lillis B, Vergona R, Post J, Sullivan ME, Verhallen PF. Reduction of cardiac functional reserve and elevation of aortic stiffness in hyperlipidemic Yucatan minipigs with systemic and coronary atherosclerosis. *Vascul Pharmacol.* 2002;39(1-2):69-76.

22. Barbeau ML, Klemp KF, Guyton JR, Rogers KA. Dietary fish oil. Influence on lesion regression in the porcine model of atherosclerosis. *Arterioscler Thromb Vasc Biol.* 1997;17(4):688-694.

23. Carter AJ, Bailey L, Devries J, Hubbard B. The effects of uncontrolled hyperglycemia on thrombosis and formation of neointima after coronary stent placement in a novel diabetic porcine model of restenosis. *Coron Artery Dis.* 2000;11(6):473-479. 24. de Smet BJ, van der Zande J, van der Helm YJ, Kuntz RE, Borst C, Post MJ. The atherosclerotic Yucatan animal model to study the arterial response after balloon angioplasty: the natural history of remodeling. *Cardiovasc Res.* 1998;39(1):224-232.

25. Gal D, Rongione AJ, Slovenkai GA, DeJesus ST, Lucas A, Fields CD, Isner JM. Atherosclerotic Yucatan microswine: an animal model with highgrade, fibrocalcific, nonfatty lesions suitable for testing catheter-based interventions. *Am Heart J.* 1990;119(2 Pt 1):291-300.

26. Dyson MC, Alloosh M, Vuchetich JP, Mokelke EA, Sturek M. Components of metabolic syndrome and coronary artery disease in female Ossabaw swine fed excess atherogenic diet. *Comp Med.* 2006;56(1):35-45.

27. Lee KT, Jarmolych J, Kim DN, Grant C, Krasney JA, Thomas WA, Bruno AM. Production of advanced coronary atherosclerosis, myocardial infarction and "sudden death" in swine. *Exp Mol Pathol.* 1971;15(2):170-190.

28. Mihaylov D, van Luyn MJ, Rakhorst G. Development of an animal model of selective coronary atherosclerosis. *Coron Artery Dis.* 2000;11(2):145-149.

29. Koktzoglou I, Harris KR, Tang R, Kane BJ, Misselwitz B, Weinmann HJ, Lu B, Nagaraj A, Roth SI, Carroll TJ, McPherson DD, Li D. Gadofluorine-enhanced magnetic resonance imaging of carotid atherosclerosis in Yucatan miniswine. *Invest Radiol.* 2006;41(3):299-304.

30. Ishii A, Vinuela F, Murayama Y, Yuki I, Nien YL, Yeh DT, Vinters HV. Swine model of carotid artery atherosclerosis: experimental induction by surgical partial ligation and dietary hypercholesterolemia. *AJNR Am J Neuroradiol.* 2006;27(9):1893-1899.

31. Keelan P, Bayes-Genis A, Kantor B, Singh R, Carlson P, Lewis D, Schwartz RS. A novel porcine model for in vivo detection of vulnerable plaque: deposition and localization of lipid-rich lesions in the coronary arterial wall [abstract]. *Circulation*. 2001;104:II-67.

32. Granada JF, Moreno PR, Burke AP, Schulz DG, Raizner AE, Kaluza GL. Endovascular needle injection of cholesteryl linoleate into the arterial wall produces complex vascular lesions identifiable by intravascular ultrasound: early development in a porcine model of vulnerable plaque. *Coron Artery Dis.* 2005;16(4):217-224.

33. Granada JF, Wallace-Bradley D, Win HK, Alviar CL, Builes A, Lev El, Barrios R, Schulz DG, Raizner AE, Kaluza GL. In vivo plaque characterization using intravascular ultrasound-virtual histology in a porcine model of complex coronary lesions. *Arterioscler Thromb Vasc Biol.* 2007;27(2):387-393.

34. Tellez A, Alviar C, Builes A, Lopez-Berestein G, Sanguino A, Wallace-Bradley D, Ballantyne C, Perrard X, Schulz D, Kleiman N, Lev E, Frangogiannis N, Kaluza G, Granada J. Pro-Inflammatory Chemokines are Expressed in the Pig Model of Complex Coronary Lesions Induced by Intramural Injection of Lipids (TCT E-Abstract 603). *Am J Cardiol.* Oct 2007;100 (suppl):232L.

35. Schwartz RS, Murphy JG, Edwards WD, Camrud AR, Vliestra RE, Holmes DR. Restenosis after balloon angioplasty. A practical proliferative model in porcine coronary arteries. *Circulation*. 1990;82(6):2190-2200.

36. Lin W, Abendschein DR, Celik A, Dolan RP, Lauffer RB, Walovitch RC, Haacke EM. Intravascular contrast agent improves magnetic resonance angiography of carotid arteries in minipigs. *J Magn Reson Imaging*. 1997;7(6):963-971.

37. Spuentrup E, Ruebben A, Mahnken A, Stuber M, Kolker C, Nguyen TH, Gunther RW, Buecker A. Artifact-free coronary magnetic resonance angiography and coronary vessel wall imaging in the presence of a new,



metallic, coronary magnetic resonance imaging stent. *Circulation*. 2005;111(8):1019-1026.

38. Bae KT, Heiken JP, Brink JA. Aortic and hepatic peak enhancement at CT: effect of contrast medium injection rate- pharmacokinetic analysis and experimental porcine model. *Radiology.* 1998;206(2):455-464.

39. Bae KT, Heiken JP, Brink JA. Aortic and hepatic contrast medium enhancement at CT. Part II. Effect of reduced cardiac output in a porcine model. *Radiology*. 1998;207(3):657-662.

40. Bae KT, Tran HQ, Heiken JP. Multiphasic injection method for uniform prolonged vascular enhancement at CT angiography: pharmacokinetic analysis and experimental porcine model. *Radiology.* 2000;216(3):872-880.

41. Rist C, Nikolaou K, Flohr T, Wintersperger BJ, Reiser MF, Becker CR. High-resolution ex vivo imaging of coronary artery stents using 64-slice computed tomography--initial experience. *Eur Radiol.* 2006;16(7):1564-1569.

42. D'Arceuil HE, de Crespigny AJ, Pelc L, Howard D, Alley M, Seri S, Hashiguchi Y, Nakatani A, Moseley ME. An MRA study of vascular stenosis in a pig model using CH3-DTPA-Gd (NMS60) and Gd-DTPA. *Magn Reson Imaging.* 2004;22(9):1243- 1248.

43. Green JD, Omary RA, Schirf BE, Tang R, Lu B, Gehl JA, Huang JJ, Carr JC, Pereles FS, Li D. Comparison of X-ray fluoroscopy and interventional magnetic resonance imaging for the assessment of coronary artery stenoses in swine. *Magn Reson Med.* 2005;54(5):1094-1099.

44. McFalls EO, Ward H, Fashingbauer P, Gimmestad G, Palmer B. Myocardial blood flow and FDG retention in acutely stunned porcine myocardium. *J Nucl Med.* 1995;36(4):637-643.

45. Chareonthaitawee P, Schaefers K, Baker CS, Turkheimer F, Stegger L, Banner NR, Yacoub M, Bonser RS, Iozzo P, Camici PG, Rimoldi O. Assessment of infarct size by positron emission tomography and [18F]2-fluoro-2-deoxy-D-glucose: a new absolute threshold technique. *Eur J Nucl Med Mol Imaging*. 2002;29(2):203-215.

46. Schafers KP, Spinks TJ, Camici PG, Bloomfield PM, Rhodes CG, Law MP, Baker CS, Rimoldi O. Absolute quantification of myocardial blood flow with H(2)(15)O and 3-dimensional PET: an experimental validation. *J Nucl Med.* 2002;43(8):1031-1040.

47. Wagner B, Anton M, Nekolla SG, Reder S, Henke J, Seidl S, Hegenloh R, Miyagawa M, Haubner R, Schwaiger M, Bengel FM. Noninvasive characterization of myocardial molecular interventions by integrated positron emission tomography and computed tomography. *J Am Coll Cardiol.* 2006;48(10):2107-2115.

48. Chareonthaitawee P, Christenson SD, Anderson JL, Kemp BJ, Hodge DO, Ritman EL, Gibbons RJ. Reproducibility of measurements of regional myocardial blood flow in a model of coronary artery disease: Comparison of H2150 and 13NH3 PET techniques. *J Nucl Med.* 2006;47(7):1193-1201.

49. Hartvig P, Waldenstrom A, Wikstrom G, Zielinski T, Martinussen HJ, Carslsten J, Voipio-Pulkki LM, Lundqvist H, Bjurling P, Nagren K, et al. The diabetic heart: a porcine model evaluated with positron emission tomography using 1-11C-palmitate and 3-11C-pyruvate. *Diabetes Res.* 1989;12(1):1-5.

50. Johnson LL, Schofield L, Donahay T, Narula N, Narula J. 99mTcannexin V imaging for in vivo detection of atherosclerotic lesions in porcine coronary arteries. *J Nucl Med.* 2005;46(7):1186-1193.

51. Vallabhajosula S, Granada JF, Kothari PJ. Molecular Imaging of Vulnerable Plaque: Comparison of 3 PET Tracers; [11C]PK11195, [18F]Fluorocholine and [18F]FDG in a Porcine Model of Complex Coronary Atherosclerosis. (Abstract). *J Nucl Med.* 2007;48:103p.

52. Mehran R, Mintz GS, Hong MK, Tio FO, Bramwell O, Brahimi A, Kent KM, Pichard AD, Satler LF, Popma JJ, Leon MB. Validation of the in vivo intravascular ultrasound measurement of in-stent neointimal hyperplasia volumes. *J Am Coll Cardiol.* 1998;32(3):794-799.

53. Asawa K, Kataoka T, Kobayashi Y, Hasegawa T, Nishioka H, Yamashita H, Qiu Z, Ehara S, Hirose M, Kamimori K, Shimada K, Yoshiyama M, Yoshikawa J. Method analysis for optimal continuous imaging using intravascular optical coherence tomography. *J Cardiol.* 2006;47(3):133-141.

54. Tearney GJ, Jang IK, Kang DH, Aretz HT, Houser SL, Brady TJ, Schlendorf K, Shishkov M, Bouma BE. Porcine coronary imaging in vivo by optical coherence tomography. *Acta Cardiol.* 2000;55(4):233-237.

55. Maeng M, Olesen PG, Emmertsen NC, Thorwest M, Nielsen TT, Kristensen BO, Falk E, Andersen HR. Time course of vascular remodeling, formation of neointima and formation of neoadventitia after angioplasty in a porcine model. *Coron Artery Dis.* 2001;12(4):285-293.

56. de Korte CL, van der Steen AF. Intravascular ultrasound elastography: an overview. *Ultrasonics*. 2002;40(1-8):859-865.

57. Hamilton AJ, Huang SL, Warnick D, Rabbat M, Kane B, Nagaraj A, Klegerman M, McPherson DD. Intravascular ultrasound molecular imaging of atheroma components in vivo. *J Am Coll Cardiol.* 2004;43(3):453-460.

58. Frohlich G, Strehblow C, Sperker W, Yahya N, Shirazi M, Hevesi A, Garamvolgyi R, Hadjiev J, Scherzer T, Glogar D, Gyongyosi M. Serial intravascular ultrasonographic measurements after implantation of biodegradable polymer-coated stents in porcine coronary arteries. *Coron Artery Dis.* 2003;14(5):409-412.

59. Zhang Q, Lu L, Pu L, Zhang R, Shen J, Zhu Z, Hu J, Yang Z, Chen Q, Shen W. Neointimal hyperplasia persists at six months after sirolimus-eluting stent implantation in diabetic porcine. *Cardiovasc Diabetol.* 2007;6:16.

